

# Anti-Diarrheal Evaluation of the Aqueous Ethanol Extract of *Jateorhiza macrantha* (Hook F.) Exell Mendonça (Menispermaceae) Leaves

Onyejekwe V. N, Afieroho OE\*, Abo KA

Department of Pharmacognosy and Phytotherapy, University of Port Harcourt, Nigeria

DOI: [10.36348/sjbr.2020.v05i02.004](https://doi.org/10.36348/sjbr.2020.v05i02.004)

| Received: 04.02.2020 | Accepted: 11.02.2020 | Published: 29.02.2020

\*Corresponding author: Afieroho OE

## Abstract

*Jateorhiza macrantha* is a medicinal plant used in ethnomedicine for dysentery, cough, boil, ulcer, and inflammation, wound healing and venomous bite. This study is aimed at evaluating the anti-diarrheal activity of the aqueous ethanol extract from the leaves of *Jateorhiza macrantha*. The aqueous ethanol extract (AEE) was obtained by exhaustive cold maceration in 70% aqueous. The anti-diarrheal and anti-motility activities of the AEE were evaluated using the castor oil induced diarrheal and charcoal meal animal assays respectively. Loperamide (2mg/kg bw) and atropine sulphate (5 mg/kg bw) were used as reference anti-diarrheal and anti-motility agents respectively for comparison. The AEE shows dose dependent anti-diarrheal and anti-motility activities. The trend in the anti-diarrheal activity (% inhibition of fecal mass compared to negative control group): loperamide-2mg/kgbw (67.42%) > AEJML-400mg/kgbw (64.49%) > AEJML-200mg/kgbw (41.57%) > AEJML-100mg/kgbw (13.48%) and anti-motility (% inhibition of intestinal peristalsis index compared to negative control group): AEJML-250mg/kgbw(71.70%) > AEJML-125mg/kgbw(30.10%) > AEJML-100mg/kgbw(0.37%). The trend in anti-castor oil induced diarrheal and anti-motility activities obtained in this study for *Jateorhiza macrantha* justifies its use in ethno medicine for the treatment of diarrheal and related gas intestinal tracts illness.

**Keywords:** *Jateorhiza macrantha*, Menispermaceae, anti-diarrheal activity, anti-motility, anti-cholinergic, drug discovery.

**Copyright © 2020:** This is an open-access article distributed under the terms of the Creative Commons Attribution license which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use (NonCommercial, or CC-BY-NC) provided the original author and source are credited.

## INTRODUCTION

Diarrhea is a disease of the gastro-intestinal tract (GIT) characterized by frequent increase of bowel movement, wet stool and abdominal pain [1]. It is a major cause of death among young babies below five years in undeveloped countries globally [2]. It has been reported that 57800 out of 6.3 million children below five years died of diarrheal related illnesses in Africa [3]. The high rate of mortality in infants due to diarrheal infection, and the high cost of orthodox medicine has resulted to the return to traditional medicine by the global population as an alternative to orthodox medicine. Medicinal plants still remain a source of novel pharmacological principle for new drugs [4]. *Jateorhiza macrantha* and its closely related species *Jateorhiza palmata* are the only two known species of medicinal plant in the genus *Jateorhiza* belonging to the Menispermaceae family- a family which also includes the medicinal and poisonous plant -St John's Wort. *J. macrantha* is widely used in ethnomedicine for the

treatment of dysentery and diarrhea, parasitic infections, miscarriage, venomous stings and bites [5-10]. It is also use in the treatment of hypertension [11], and dysmenorrheal [12], weak libido, wound and ulcer [9, 10]. Chemical compounds like: columbin, isocolumbin, plamarin, isojectorin, chasmanthin and palmatosides A are reported found in its closely related species *Jateorhiza palmata* [13] with scarce literature on similar reports for its closely related species *Jateorhiza palmata* [13] with scarce literature on similar reports for *Jateorhiza macrantha*. This study is aimed at evaluating the activity guided antidiarrheal fractionation of the aqueous ethanol extract of the leaves of *Jateorhiza macrantha* as a preliminary step toward the isolation and structural elucidation its anti-diarrheal constituent(s) that could serve as useful leads in drug development.

## MATERIALS

Analytical grade reagents and chemicals used were products of BDH and/ or Sigma Aldrich unless otherwise stated. Loperamide hydrochloride was bought from Boldstep pharmaceutical shop Port-Harcourt, Rivers State Nigeria. Atropine sulphate (Sigma-Aldrich). The fresh leaves of *Jateorhiza macrantha* were sourced from Ijebu forests in Ore Ondo State, South Western Nigeria and authenticated by a Taxonomist Dr. Chukwuka S. Kanayo of the Department of Botany University of Ibadan with Voucher specimen number UIH 22610 deposited in the herbarium of same University. The leaves were air dried at room temperature, ground to fine powder and stored in an air tight container for further use. Swiss albino mice of both sex (6-7 weeks old) which weighed between 25-30g, and wistar albino rats of both sex (150-200 g) used in this study were collected from Department of Pharmacology, University of Port-Harcourt, Nigeria. The animals were maintained under standard environmental conditions for acclimation for at least 1 week in the animal house before experiment. They had free access to food, and water ad libitum. The study protocol was approved by the institutional animal ethical Committee University of Port-Harcourt, Rivers State Nigeria (UPH/CEREMAD/REC/MM/60/036).

## METHODS

### Preparation of the aqueous ethanol extract of *Jateorhiza macrantha* leaves

This was done following a prior protocol established in our laboratory. Briefly, the aqueous ethanol extract (AEE) was obtained by exhaustive cold maceration of the powdered *Jateorhiza macrantha* leaves (300 g) at room temperature in 70% ethanol for 72 hours. The extraction solvent was replaced with fresh solvent at 24 hours interval during the 72 hours period of extraction. The filtrates obtained were pooled and concentrated using a rotary evaporator and further dried over the water bath at < 45°C to afford the crude 70 % aqueous ethanol extract (AEE).

### Phytochemical Screening

This was done using standard phytochemical screening reagents for the presence of the plant metabolites: alkaloids, saponins, tannins, flavonoids, phlobatannins, cyanogenic glycosides, cardiac glycosides, anthraquinones, triterpenoids, sugars, as reported [14, 15].

### Castor oil induced Diarrhea in mice

This test was done according to the reported method [16, 17] with slight modification. Briefly, thirty mice were fasted for 18 – 24 hours and divided into five groups with six animals in each group. First group received distilled water 10ml/kg bw and the second received loperamide (2mg/kg), serving as negative and positive respectively. Group 3, 4 and 5 received 100, 200 and 400mg/kg of the extract respectively. After 45minutes, the animals received separately, 0.3ml of

castor oil orally. The animals were kept in separate cages underplayed with a pre-weighed white paper. The severity of diarrhea was observed for 4hours. The mean weight of pasty liquid or wet defecation (diarrheal droppings) and time of onset of diarrhea was determined and compared with the negative control group. The total score of diarrheal feces for the negative control group was considered as 100% non-inhibition effect. The percentage inhibition of diarrhea was calculated using the formula:

$$\% \text{ inhibition of diarrhea} = \frac{\text{MWDC} - \text{MWDT}}{\text{MWDC}} \times 100$$

Where,

MWDC is the mean weight of wet defecation due to diarrhea (negative) control group.

MWDT is the mean weight of wet defecation due to test group.

### Charcoal meal gastro-intestinal motility test

The effect of the extract on small intestinal transit was studied in overnight fasted wistar albino rat (150- 200 g) of either sex, which were divided into six groups containing four rats in each group including positive and negative controls. The negative control was given 2ml of DMSO while the positive control was given 10 mg/kg atropine sulphate orally. Other groups were given orally 100, 125, and 250 mg/kg body weight respectively. Five minutes after treatment, all rats were given 0.5 ml of 5% charcoal in 5% gum acacia by oral route. All animals were sacrificed after 30 mins, the intestine removed, and the mean total length of the small intestine (TLSI), and the mean transit distance by the charcoal meal in the small intestine (TDCM) were measured. The peristaltic index (PI) and the percentage inhibition of intestinal motility were calculated as shown below [17, 18].

$$\text{Peristaltic index (PI)} = \frac{\text{TDCM} \times 100}{\text{TLSI}}$$

$$\% \text{ Inhibition of intestinal motility} = \frac{\text{PIC} - \text{PIT}}{\text{PIC}} \times 100$$

Where,

TLSI is the mean total length of the small intestine.

TDCM is the mean distance covered by the charcoal meal in the small intestine.

PIC is the peristaltic index due to diarrhea (negative) control group.

PIT is the peristaltic index due to test group.

## STATISTICAL ANALYSIS

All results are expressed as mean  $\pm$  standard error of mean (SEM). All statistical analyses were performed by one- way ANOVA followed by the

Tukey HSD post-hoc test where  $P < 0.05$  was regarded as statistical significant.

## RESULTS AND DISCUSSION

The 70% aqueous ethanol extract obtained by cold maceration yield 3.98% w/w. From the phytochemical screening of the powdered leaves of *J. macrantha*, it was observed that whereas secondary plant metabolites like: saponins, triterpenoids, cyanogenic glycosides, cardiac glycosides, and anthraquinones were absent, phenolic constituents like tannins and flavonoids, carbohydrate derivatives in form of free and combined reducing sugars and alkaloids are evidently present. The observed trend in the phytochemical constituents is in agreement with that reported in the literature [10]. The crude 70 % aqueous ethanol extract of *J. macrantha* (AEJML) significantly ( $p < 0.05$ ) decrease in a dose dependent manner, the diarrheal fecal mass in the castor oil induced diarrheal mice as shown in Table-1. At the highest dose of 400 mg/kg bw, the anti-diarrheal effect, though slightly lower was not significantly different ( $p > 0.05$ ) compared to that due to the reference drug loperamide at 2 mg/kg bw. At this highest dose, the AEJML also relatively delayed the onset of diarrheal episode more than the reference drug loperamide at 2 mg/kg bw though this effect was not significant ( $p > 0.05$ ). Thus, at the highest dose of 400 mg/kg bw the antidiarrheal activity perception or index of the AEJML could be seen as being comparable with the reference drug loperamide (2 mg/kg bw). Similar trend was also observed for the charcoal meal gastro- intestinal anti-motility test. Here, all the test AEJML doses and the reference drug loperamide significantly inhibited the charcoal meal intestinal transit time compared to the control group as seen in Table-2 with a trend in the peristaltic index (PI) of: AEJML 250 mg/kgbw (PI=24.79 %) < loperamide 2 mg/kg bw (PI= 35.29 %) < AEJML 125 mg/kgbw (PI=61.23 %) < AEJML 100 mg/kgbw (PI=87.28 %) < negative control group (PI = 87.60 %). This trend is further buttressed by that for the percentage inhibition of GIT motility (see also Table-2) with the trend: AEJML 250 mg/kgbw (71.70 %) > atropine sulphate 10

mg/kg bw (59.71 %) > AEJML 125 mg/kgbw (30.10 %) > AEJML 100 mg/kgbw (PI=0.37 %). Thus, the AEJML could be seen as inhibiting both castor oil induced secretory diarrhea and gastro-intestinal motility or peristalsis. Depending on the cause, diarrhea may be marked by increased electrolyte secretion (secretory diarrhea), increased luminal osmolality (osmotic diarrhea), decreased electrolyte absorption and/or increased intestinal motility [17]. In line with this, the mechanism of action of any given anti-diarrheal agent could be by increasing the GIT transit time by way of inhibiting GIT motility as seen in the charcoal meal test or by inhibition of intestinal secretions and/or increasing the intestinal absorption of water and electrolytes [17]. Castor oil is a laxative due to its containing ricinoleic acid as the laxative principle which has been found to inhibit intestinal  $\text{Na}^+/\text{K}^+$  ATPase activity as well as stimulating the biosynthesis and release of endogenous prostaglandins responsible for diarrhea [17, 19, 20]. Several plant extracts and plant derived metabolites have been shown to exhibit anti-diarrhea activity by inhibiting castor oil induced secretory diarrhea or by the anticholinergic mechanism as seen in the charcoal meal anti-motility assay model [16-20]. Plant secondary metabolites like alkaloids, and phenolics like flavonoids, tannins and phenylpropanoids have been reported to exhibit anti-diarrhea activity. Tannins and flavonoids are phenolic compounds occurring naturally in plants with ecological function. Flavonoids inhibit the release of prostaglandin by altering the production of lipooxygenase and cyclooxygenase I and II [21]. Thus flavonoids control the secretion induced by castor oil and make electrolytes absorption easier [22]. It was also reported that flavonoids [23] and tannin [24] have antidiarrheal property because they are able to inhibit the motility of the intestine. Therefore the presence of flavonoids, tannins and alkaloids in the leaves of *J. macrantha* in this study, offer a plausible rationale for the observed trend in the inhibitory effect on castor oil induced diarrhea and charcoal meal gastro intestinal motility.

**Table-1: Effect of *Jateorhiza macrantha* Leaves 70 % aqueous ethanol extract on Castor Oil-induced diarrhea in wistar albino mice**

Test Groups	Time of diarrheal onset (Mins)	Weight (g) of fecal mass	% Diarrheal inhibition
Negative Control (10ml/kg bw)	20.50 ± 1.70	4.45 ± 0.66	-
Loperamide (2 mg/kg)	28.75 ± 5.85	1.45 <sup>a</sup> ± 0.31	67.42
AEJML (100 mg/kg)	23.50 ± 2.65	3.85 ± 0.47	13.48
AEJML (200 mg/kg)	27.75 ± 5.32	2.60 ± 0.39	41.57
AEJML (400 mg/kg)	32.55 ± 5.92	1.58 <sup>a</sup> ± 0.30	64.49

Key: AEJML 70 % aqueous ethanol extract of *J. macrantha* leaves, values are Mean ± SEM, <sup>a</sup> represent the values significantly difference  $P < 0.05$  from the negative control.

**Table-2: Effect of the *Jateorhiza macrantha* Leaves 70 % aqueous ethanol extract on charcoal meal intestinal transit time in wistar albino rats**

Groups	Length of small intestine (cm)	Distance moved by the charcoal meal(cm)	Peristaltic index PI	Percentage inhibition of GIT motility
Negative control (10 ml/kg bw)	42.81 ± 3.78	37.5 ± 3.54	87.60	-
Atropine(10 mg/kg bw)	22.50 ± 2.74	7.94 <sup>a</sup> ± 1.63	35.29	59.71
AEJML(100 mg/kg bw)	22.56 ± 2.74	19.69 <sup>a</sup> ± 2.56	87.28	0.37
AEJML(125 mg/kg bw)	19.19 ± 2.53	11.75 <sup>a</sup> ± 1.98	61.23	30.10
AEJML(250 mg/kg bw)	29.25 ± 3.12	7.25 <sup>a</sup> ± 1.56	24.79	71.70

Key: AEJML 70 % aqueous ethanol extract of *J. macrantha* leaves, values are Mean ± SEM, <sup>a</sup> represent the values significantly difference  $P < 0.05$ ) from the negative control.

## CONCLUSION

The observed anti- castor oil induced diarrhea and charcoal meal transit time inhibition effects of the aqueous ethanol extract of *J. macrantha* in this study, and the presence of flavonoids, tannins and alkaloids which are bioactive secondary metabolites reported to have anti-diarrhea and anti-motility activities have confirmed scientifically the use of this plant in ethnomedicine for the treatment and management of diarrhea and related GIT illnesses. Further work is ongoing to isolate and elucidate the chemical structure of the pure form of the constituents in the aqueous ethanol extract of *J. macrantha* using spectroscopy techniques, and evaluate them for possible development into anti-diarrheal medications for clinical use.

## REFERENCES

- Maiti, A. (2007). In Vivo Evaluation of Antidiarrheal Activity of the Seed of Swietenia Tropical. *Journal Pharmaceutical Research*. 6(2):711-716.
- Fischer-Walker, C. L., Rudan, I., Liu, L., Nair, H., Theodoratou, E., Bhutta, Z. A., O'Brien, K. L., Campbell, H., & Black, R. E. (2013). Global burden of childhood pneumonia and diarrhea. *Lancet* 381: 1405-1416.
- Liu, L., Johnson, H. L., Cousens, S., Perin, J., Scott, S., Lawn, J. E., ... & Mathers, C. (2012). Child Health Epidemiology Reference Group of WHO and UNICEF Global, regional, and national causes of child mortality: an updated systematic analysis for 2010 with time trends since 2000. *Lancet*, 379(9832), 2151-2161.
- Gupta, P. D., & Birdi, T. J. (2017). Development of botanicals to combat antibiotic resistance. *Journal of Ayurveda and integrative medicine*, 8(4), 266-275.
- Dalziel, J. M. (1937). The useful plants of W. Tropical Africa, Crown agents for the colonies, London. 18-22.
- Burkill, H. M. (1985). The useful plants of W. Tropical Africa. Royal Botanical Gardens, Kew. 4, 143.
- Irvine, J. R. (1961). Woody Plants of Ghana. Oxford University press, London. 32-34.
- Oliver, B. (1960). Medicinal plants in Nigeria. College of Arts, Science and Technology, Lagos. 139.
- Ajayi, G. O., Salako, O. A., & Mosebolatan, M. I. (2015). Anti-inflammatory and Analgesic activities of *Jateorrhiza macrantha* (Menispermaceae). *International Journal of phytomedicine*, 7(4):427-431.
- Ajayi, G. O., Kadiri A. B., & Ibiloye A. E. (2015). Pharmacognostic Characteristics of the West African *Jateorhiza macrantha* (Hook.F.) Exell & Mendonça (Menispermaceae). *International Journal of Pharmacognosy and Phytochemical Research*, 7(5):1001-1006.
- Aboubakar, O. B. F., Bella, N. M. T., Ngo, L. T. E., Bilanda, D. C., & Dimo, T. (2012). Antihypertensive activity of *Jateorhiza macrantha* (Menispermaceae) aqueous extract on ethanol-induced hypertension in wistar. *Int J Pharm Pharm Sci*, 4, 293-298.
- Jiofack, T., Fokunang, C., Guedje, N., Kemeuze, V., Fongzossie, E., Nkongmeneck, B. A., ... & Tsabang, N. (2010). Ethnobotanical uses of medicinal plants of two ethnobotanical regions of Cameroon. *International Journal of Medicine and Medical Sciences*, 2(3), 60-79.
- Yonemitsu, M., Fukuda, N., Kimura, T., & Komori, T. (1987). Studies on the Constituents of *Jateorhiza palmata* Miers (Colombo Root), II. Separation and Structure of Six New Furanoid Diterpene Glucosides: Palmatoside B, C, D, E, F, and G. *Liebigs Annalen der Chemie*, 1987(3), 193-197.
- Houghton, P. J., & Raman, A. (1999). Laboratory handbook for the fractionation of natural extracts. Chapman and Hall, London, UK.

15. Harborne, J. B. (1998). *Phytochemical methods-a guide to modern techniques of plant analysis*. 3rd edition, Chapman and Hall, London, UK, 302.
16. Awouter, F., Neimegeers, C. J. E., Lenaeri, F. M., & Janssen, P. A. J. (1978). Delay of castor oil diarrhoea in rats; A new way to evaluate inhibitors of prostaglandin's biosynthesis. *Journal of Pharmaceutical Pharmacology*; 30: 41-45
17. Tagne, M. A. F., Rékabi, Y., Noubissi P. A., Fankem, G. O., Akaou, H., Wambe, H., & Kamgang, R. (2019). Evaluation of Antidiarrheal Activity of Aqueous Leaf Extract of *Anogeissus leiocarpus* on Castor Oil-Induced Diarrhea in Rats. *American Journal of Biomedical Science & Research*, 3(1):1-8.
18. Pazhani, G. P., Subramanian, N., Arunchalam, G., Hemalatha, S., & Ravichandran, V. (2001). Antidiarrheal potential of *Elephantopus scaber* Linn leaf extract. *Ind drugs*, 38(5): 269-271.
19. Rawat, P., Singh, P. K., & Kumar, V. (2017). Evidence based traditional anti-diarrheal medicinal plants and their phytocompounds. *Biomedicine and Pharmacotherapy*; 96: 1453-1464.
20. Tadasse, W. T., Hailu A. E., Gurmu, A. E., & Mechesso, A. F. (2014). Experimental assessment of antidiarrheal and antiscratory activity of 80% methanolic leaf extract of *Zehneria scabra* in mice. *BMC Complementary and Alternative Medicine*; 14(1):460-468.
21. Awad, A. B., Toczek, J., & Fink, C. S. (2004). Phytosterols decrease prostaglandin release in cultured P388D1/MAB macrophages. *Prostaglandin Leukotrienes & Essential Fatty acid*, 70(6): 511-520.
22. Hamalainen, M., Nieminen, R., Asmaw, M. Z., Vuorela, P., Vapaatalo, H., & Moilanen, E. (2011). Effects of Flavonoids on Prostaglandin E2 Production and on COX-2 and mPGES-1 Expressions in Activated Macrophages. *Planta Medica*, 77(13):1504-1511.
23. Di Carlo, G., Autore, G., Izzo, A. A., Maiolino, P., Mascolo, N., Viola, P., ... & Capasso, F. (1993). Inhibition of intestinal motility and secretion by flavonoids in mice and rats: structure- activity relationships. *Journal of Pharmacy and Pharmacology*, 45(12), 1054-1059.
24. Ashok, P. K., & Upadhyaya, K. (2012). Tannins are astringent. *Journal of pharmacognosy and phytochemistry*, 1(3), 45-50.